



## **Improved Cleaning Methods for Planetary Protection Bioburden Reduction**

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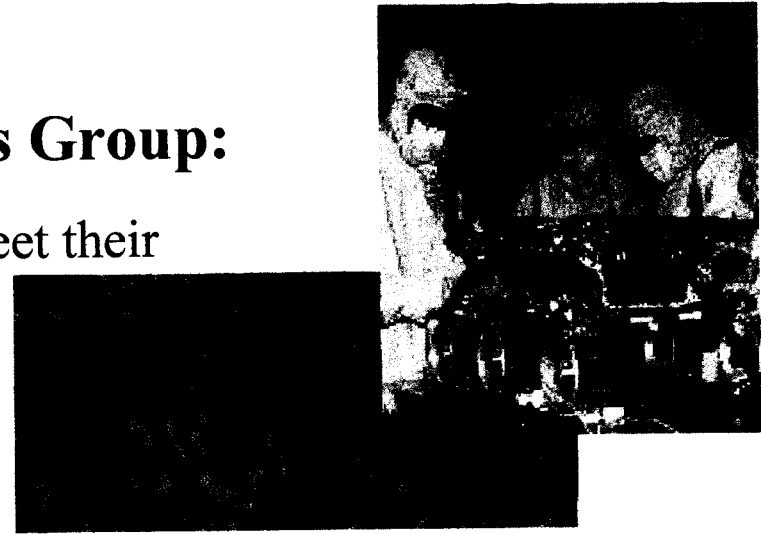
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# Introduction

## JPL Planetary Protection Technologies Group:

- Develop technologies to enable missions to meet their planetary protection requirements.
- Support science objectives of life detection missions.



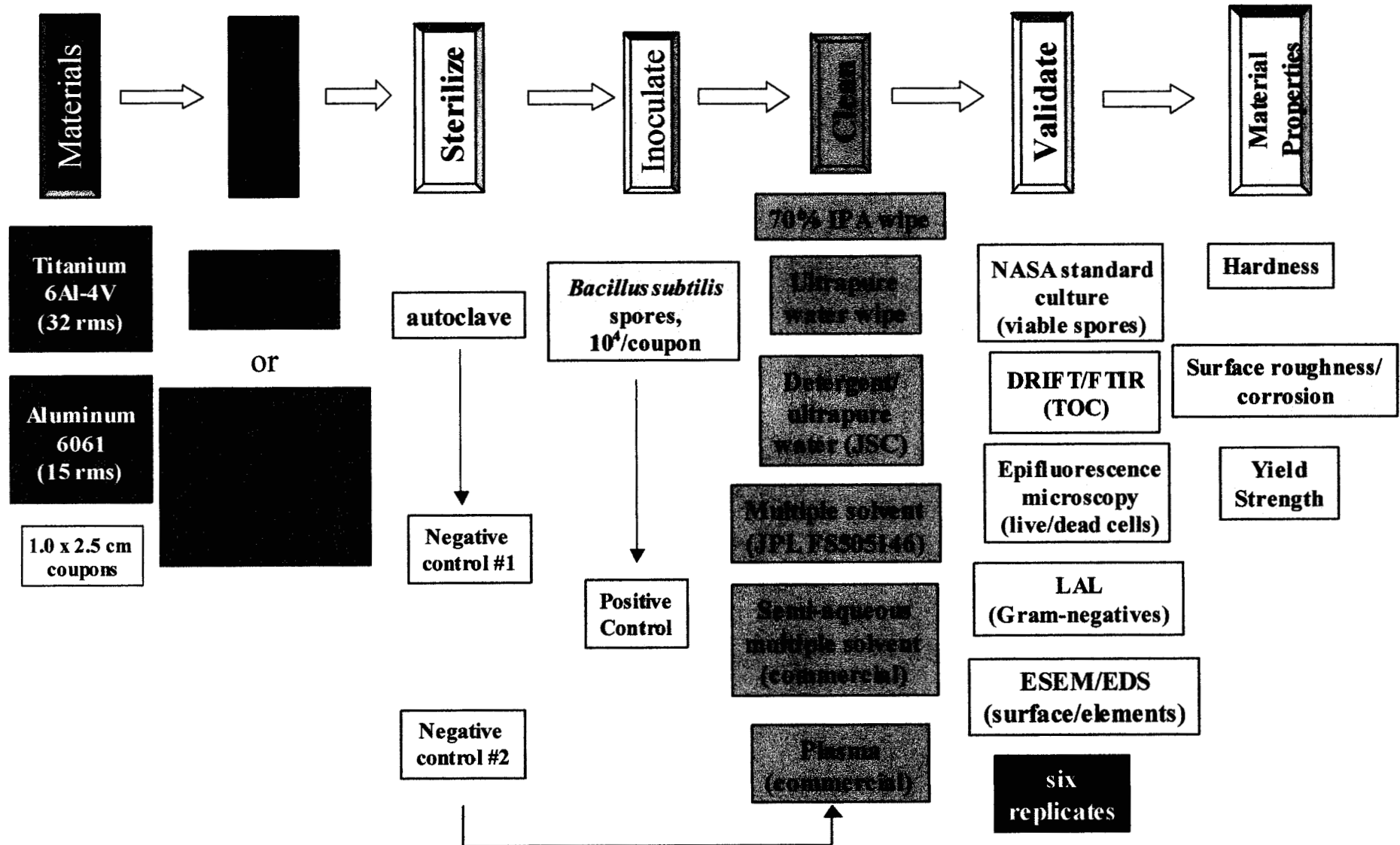
## Planetary Protection Technologies:

- **Clean** - remove biological contamination, i.e. particles, viable/nonviable organisms and residues from hardware.
- **Sterilize** - eliminate viable organisms on hardware.
- **Validate** - determine hardware biological decontamination effectiveness by quantifying and identifying remaining biological contamination.
- **Archive** - record biological history of hardware materials, assembly area, spacecraft, and launch site against future PP and science requirements.
- **Maintain** – develop cross contamination modeling tools and develop methods to identify/minimize/ remove any cross contamination or recontamination during assembly, transit, functional hardware test, rework, landing and any Earth return.

## Cleaning Technologies:

- Designed a comprehensive matrix to study the effectiveness of a variety of methods for biologically cleaning space flight hardware materials, i.e.
  - sample handling or in-situ experiment hardware materials
  - general (bulk) hardware materials
- Matrix consists of coupons of hardware materials (identified by Mars Sample Return Project) inoculated with *Bacillus subtilis* spores.
- *Bacillus subtilis* spores are NASA “standard bug,” are hardy and difficult to clean
- Coupons were then cleaned using a variety of cleaning methods and the degree of biological cleanliness is measured using a suite of techniques.

# Cleaning Matrix




# Experimental

## Materials

1.0 x 2.5 cm  
coupons

- **Aluminum, Al 6061 T6** (mill finish 15 rms) & mirror polished (2 rms, limited matrix) (0.65 Si, 0.44 Fe, 0.27 Cu, 0.02 Mn, 0.96 Mg, 0.20 Cr, 0.02 Ti)
- **Titanium, Ti 6Al 4V** (mill finish 32 rms) (0.02 C, 0.152 O, 0.15 Fe, 0.015 N, 6.4 Al, 3.9 V)

- 
- coupons placed, in a single layer, into pyrex petri dishes
  - covered petri dishes were placed in steam autoclave bags and sterilized at 121°C for 15 minutes, followed by a 30-minute drying cycle

## Sterilization

- clean room polyester wipes saturated with acetone to remove residual adhesive;
- freon vapor degreased for one hour then rinsed with isopropyl alcohol and dried

Negative  
Control #1

- no further treatment

Negative  
Control #2

- cleaned, but NOT inoculated

## Inoculation

- coupons were inoculated with 100  $\mu$ l water suspension of  $5.8 \times 10^3$  culturable ( $1.4 \times 10^4$  total) *Bacillus subtilis* spores in water and allowed to dry in air

**Positive  
Control**

- inoculated, but NOT cleaned

## Cleaning Methods

### 70% Isopropyl Alcohol (IPA) Wipe

- recommended JPL cleaning method
- mechanically wiped with 9"x9" clean room polyester wipes (Coventry 6209 c-prime, freon washed) saturated with a 70% v/v IPA/ultrapure water; performed in Class 100 biohazard hood or Class 100 laminar flow bench wearing powder-free nitrile gloves

### Ultrapure Water (UPW) Wipe

- previous studies found that some vegetative cells are better removed with pure water than with 70% IPA
- mechanically wiped with 9"x9" clean room polyester wipes (Coventry 6209 c-prime, freon washed) saturated with certified Sigma (Cat. No. W-4502) 18 M $\Omega$  water; performed in Class 100 biohazard hood or Class 100 laminar flow bench wearing powder-free nitrile gloves

## **Multiple Solvent**

- JPL Manufacturing Process Specifications FS505146 Rev. C recommended method for cleaning Al and Ti flight hardware materials
- involves ultrasonic cleaning with acetone and IPA, followed by alkaline cleaning with Oakite 61B with deionized water rinse and drying with clean, dry nitrogen; for Ti *only* procedure includes nitric acid passivation step after alkaline cleaning.
- procedure validated using MIL-STD 1246C (UV test at 3000-4000 nm, particle counts, molecular contamination)

## **Detergent/ultrapure water (UPW)**

- employed by Lockheed Martin/JSC Curation Team
- rub surfaces with dilute Joy detergent using soft, polyester knit cloth and rinse with UPW; repeat rub/rinse 3 times
- agitate coupons in nitrogen-agitated UPW bath at 75°C for 30 minutes; then air dry (d

## **Semi-aqueous, multiple solvent**

- commercially available, integrated cleaning method
- same method used for both Al and Ti
- acid cleaner for removing bacteria followed by deoxygenated metal cleaning solution then rinse with 18 M $\Omega$ , degassed, deionized water and dried with hot, clean nitrogen at 74-82°C

## **Plasma cleaning**

- commercially available, oxygen-plasma cleaning method

## Validation

### NASA Standard Cultures

- as per NASA Procedure for the Microbial Examination of Space Hardware (NPG: 5340.1C)

#### Three culturing procedures performed:

- |   |   |   |
|---|---|---|
| • Sterility assay: culture entire coupon in (TSB, tryptic soy broth)                            | ➡ | detects any culturable spores remaining on coupon after cleaning        |
| • Supernatant assay: sonicate coupon then culture supernatant on a TSA (tryptic soy agar) plate | ➡ | culturable spores removed from coupon after cleaning and sonication     |
| • Coupon assay: culture sonicated coupon on TSA plate   | ➡ | culturable spores NOT removed from coupon after cleaning and sonication |

### DRIFT/FTIR

- method used for monitoring hardware molecular contamination, per MIL-STD 1246C
- organic contamination is extracted from the surface of the coupon with dichloromethane, and the residue is evaporated onto KBr power under dry nitrogen
- performed on coupons and spore inoculant solution



## **Epifluorescence microscopy**

- method front-lined as new flight hardware cleaning validation technique
- coupons are dipped in solution of Molecular Probe Syto-9 dye and then counted
- also washed with Triton-X, collected onto a 25mm filter and counted

## **Limulus Amebocyte Lysate (LAL)**

- highly specific and sensitive assay for Gram-negative bacteria that makes use of the unique immune system enzyme cascade initiated in the blood cells (amebocytes) of *Limulus polyphemus* (horseshoe crab)
- conducted by Marine Biological Laboratory, Woods Hole

## **Environmental Scanning Electron Microscopy/Energy Dispersive Spectroscopy (ESEM/EDS)**

- ESEM allows field-of-view down to  $\mu\text{m}$  level
- EDS provides elemental analysis of  $\mu\text{m}$  sized surface features

### **Materials Properties**

- surface roughness/corrosion
- yield strength
- hardness

# Results

Material	Positive control	70% IPA	UPW	Detergent/UPW	Multiple solvent*	Semi-aqueous multiple solvent
Al 6061 (15 rms)	Y(1378)	Y Y(8) Y	Y Y(8) Y	Y Y(4) Y	Y Y(3) Y	Y Y(849) Y
	200	140	800	430	100	60
Ti 6Al-4V (32 rms)	Y(2950)	Y Y(106) Y	Y Y(98) Y	Y Y(19) Y	N N(1) N	N N(1) N biological remnants seen
	130	110	1500	1400	51	100

## • NASA Standard Culture

- sterility assay (Y/N)
- supernatant assay (culture count)
- coupon assay (Y/N)

## • DRIFT/FTIR (ng/cm<sup>2</sup>)

- results from 6 pooled coupons
- LOD = 50 ng/cm<sup>2</sup>

## • Secondary contamination

- Agar plate (culture count)
- LAL (EU/coupon)

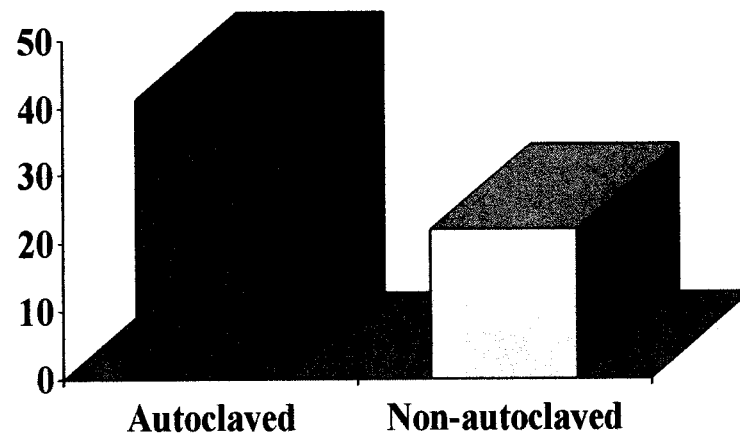
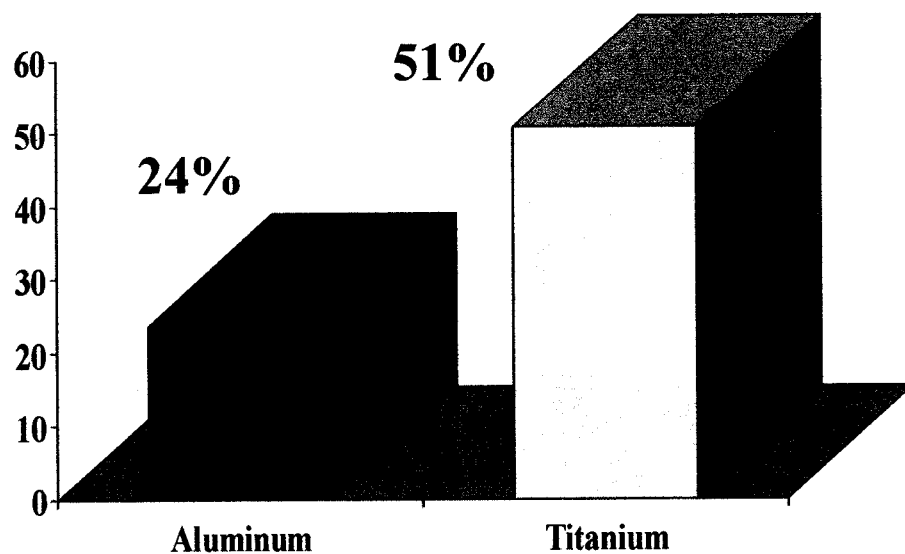
\*validated per MIL-STD 1246C at particle cleanliness level of 100

\*non pre-cleaned positive and negative controls

\*only tested positive

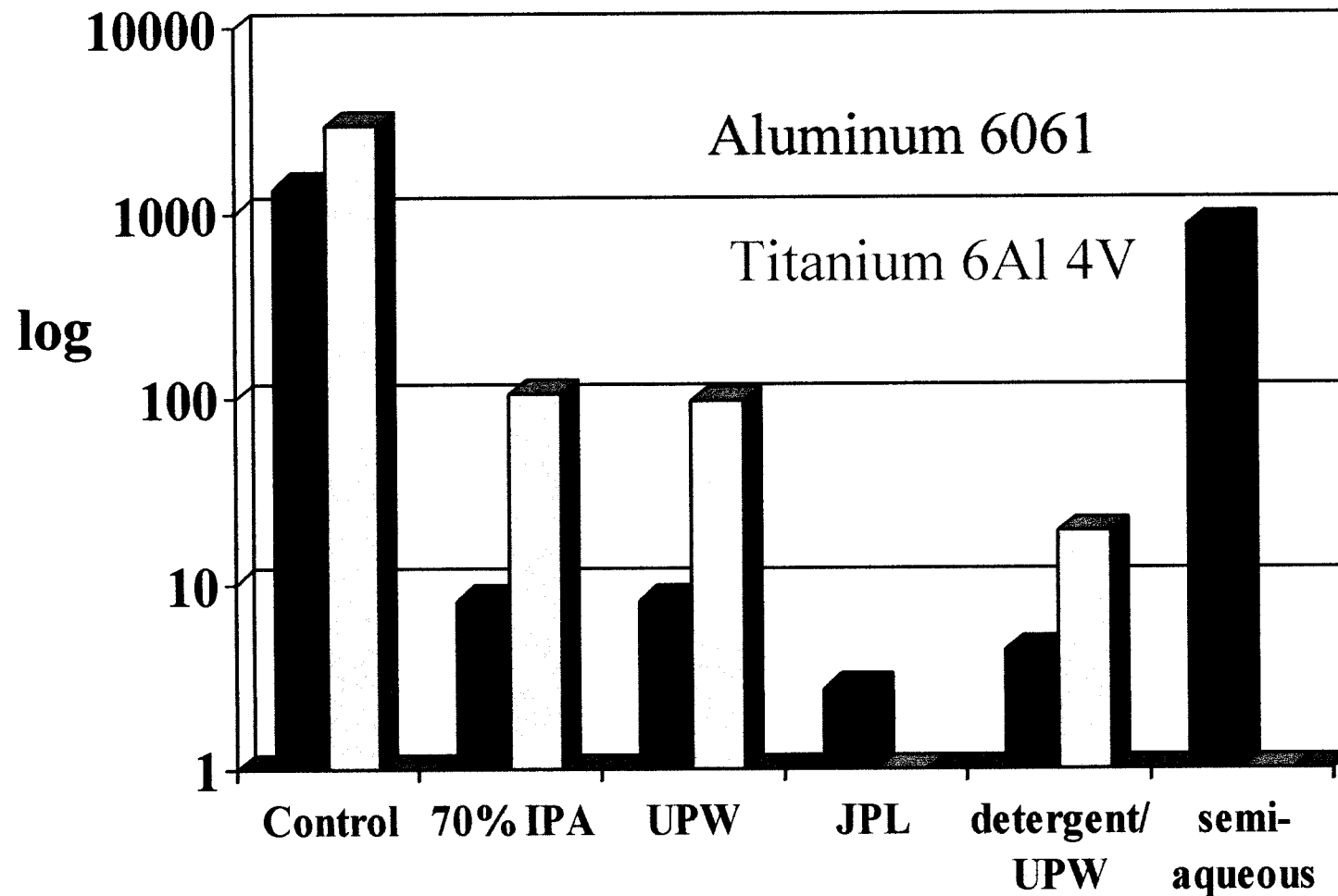
# Supernatant Assay

## % of recoverable culturable spores in supernatant

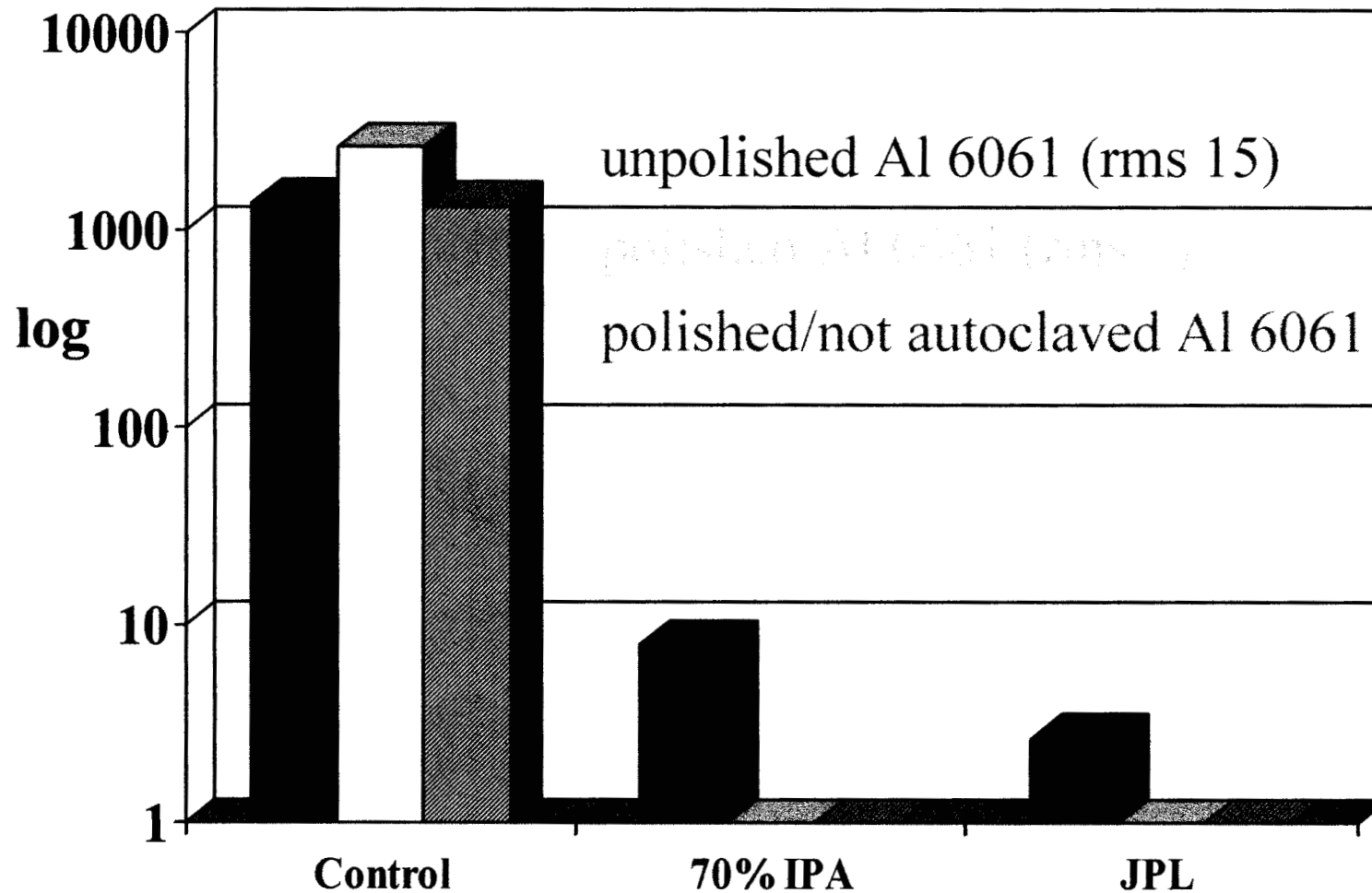


## Supernatant Assay

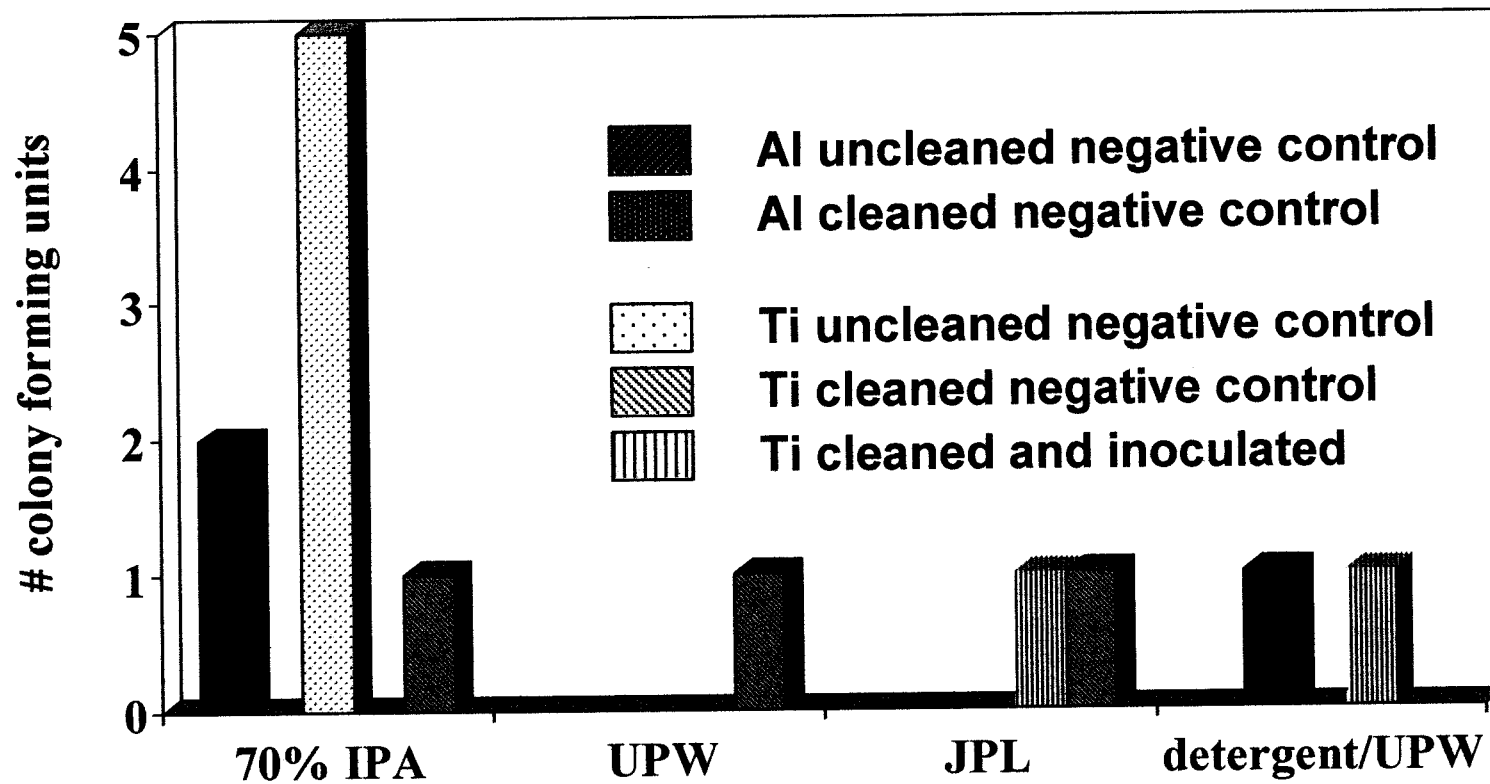
*Uncorrected!!!* # of recoverable culturable  
spores in supernatant



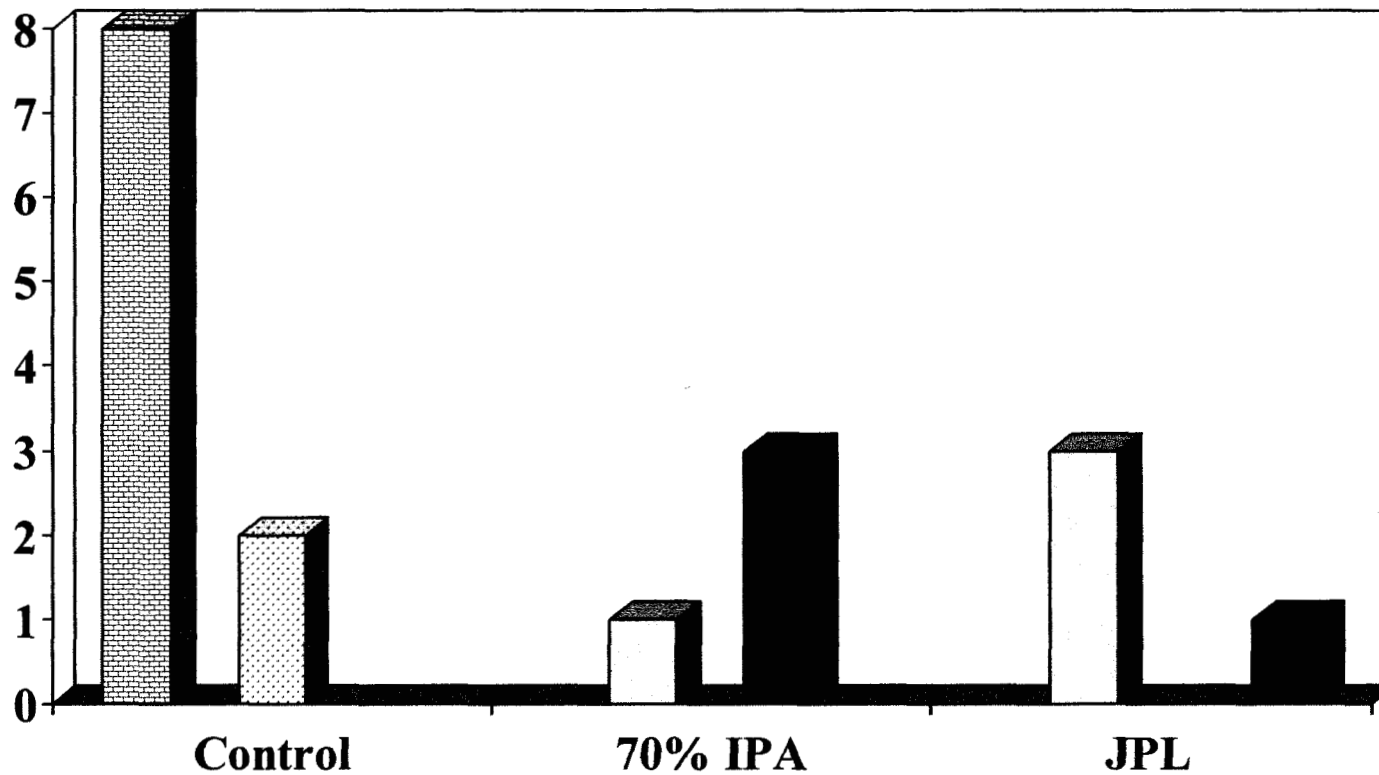
**Supernatant Assay**  
**Preliminary Results Polished Al**  
# of recoverable culturable spores in supernatant



## Coupons with secondary contamination unpolished Al 6061 and Ti 6Al 4V



## Coupons with secondary contamination polished Al 6061 (2 rms)



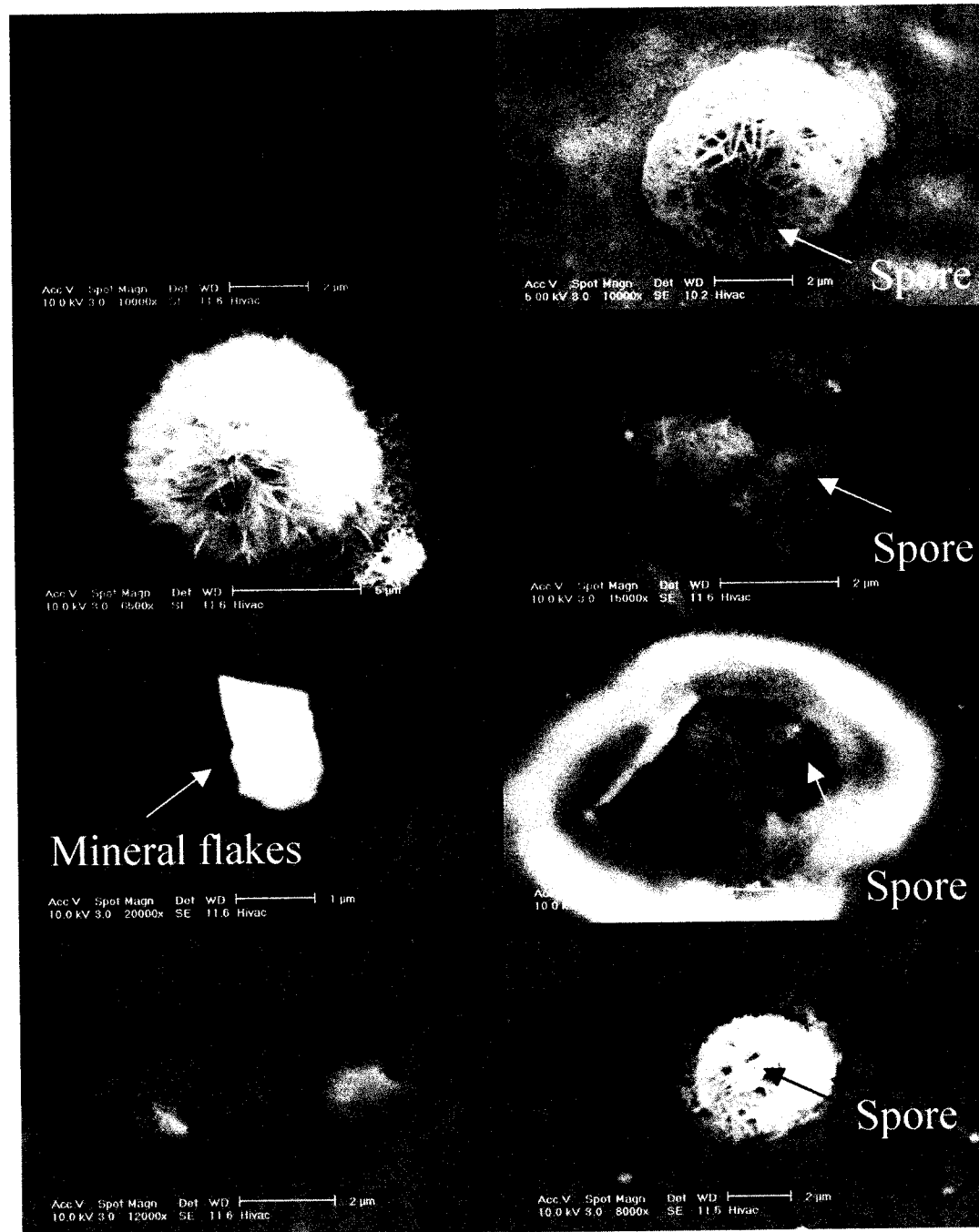
**PA: Polished & sterilized aluminum; PUS: Polished non-sterilized aluminum**

- sources of secondary biological contamination are low for unpolished Al 6061 and Ti 6Al 4V studies.
- secondary, non-biological, organic contamination measured with DRIFT/FTIR for unpolished Al 6061 and Ti 6Al 4V consists of traced aliphatic esters (commonly seen on cleaned flight hardware), silicone, organic acids, and aliphatic hydrocarbons traced to clean room wipes used, possible incomplete pre-cleaning of coupons and aluminum foil used to cover beakers.
- for polished Al 6061, secondary biological contamination also low, with LAL assays all negative, or low.
- secondary organic contamination eliminated in polished Al study, i.e. residual organic contamination found to be  $<50 \text{ ng/cm}^2$  DRIFT/FTIR limit of detection.



**Aluminum  
6061  
unpolished  
(15 rms)**

**ESEM**



control

multiple solvent

detergent/  
ultrapure water

semi-aqueous  
multiple solvent

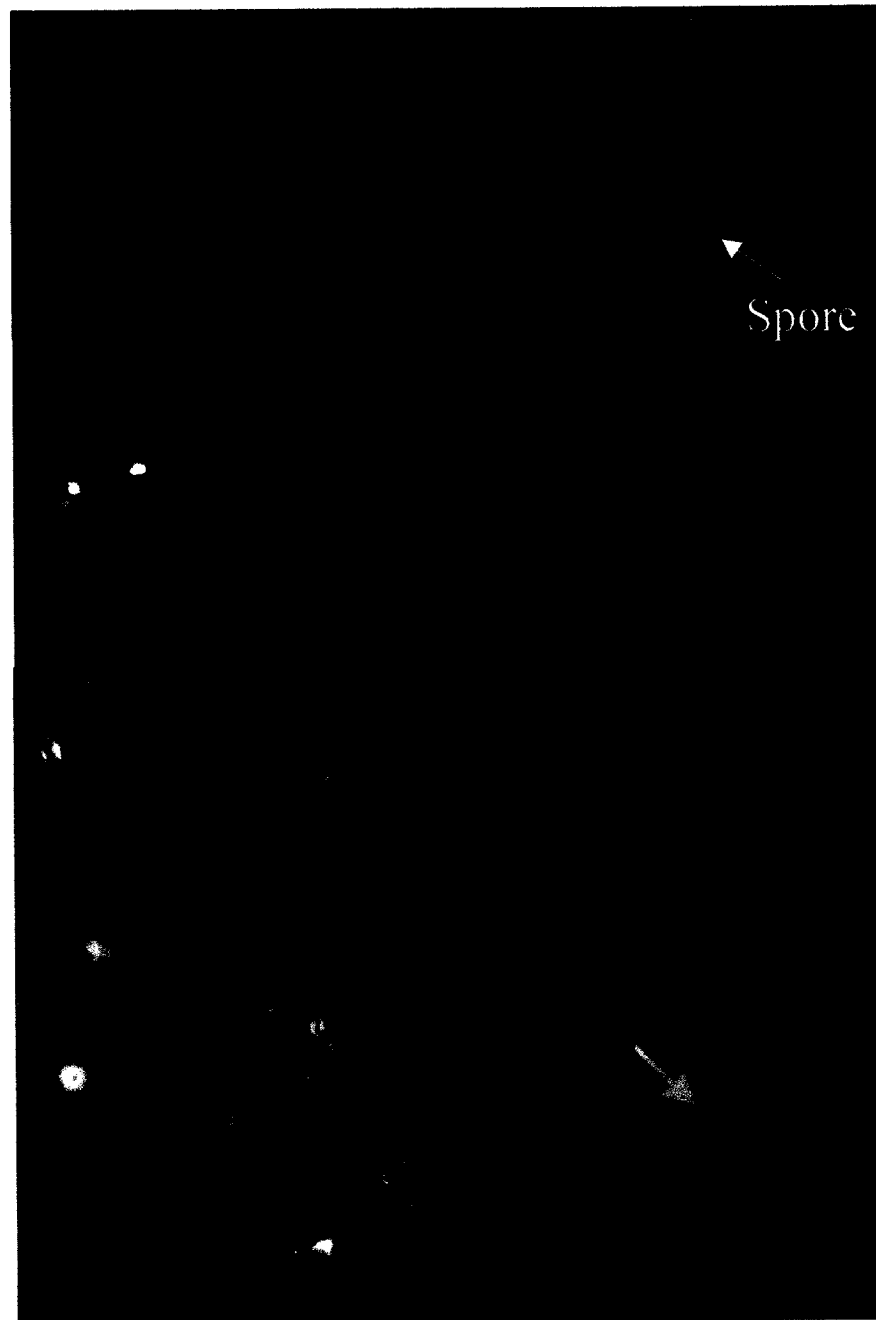
not inoculated

inoculated

**Aluminum  
6061  
unpolished  
(15 rms)**

epifluorescence  
microscopy

vegetative cell  
germinated  
from *B. subtilis*  
spores →



coupon surface

filter collected

control

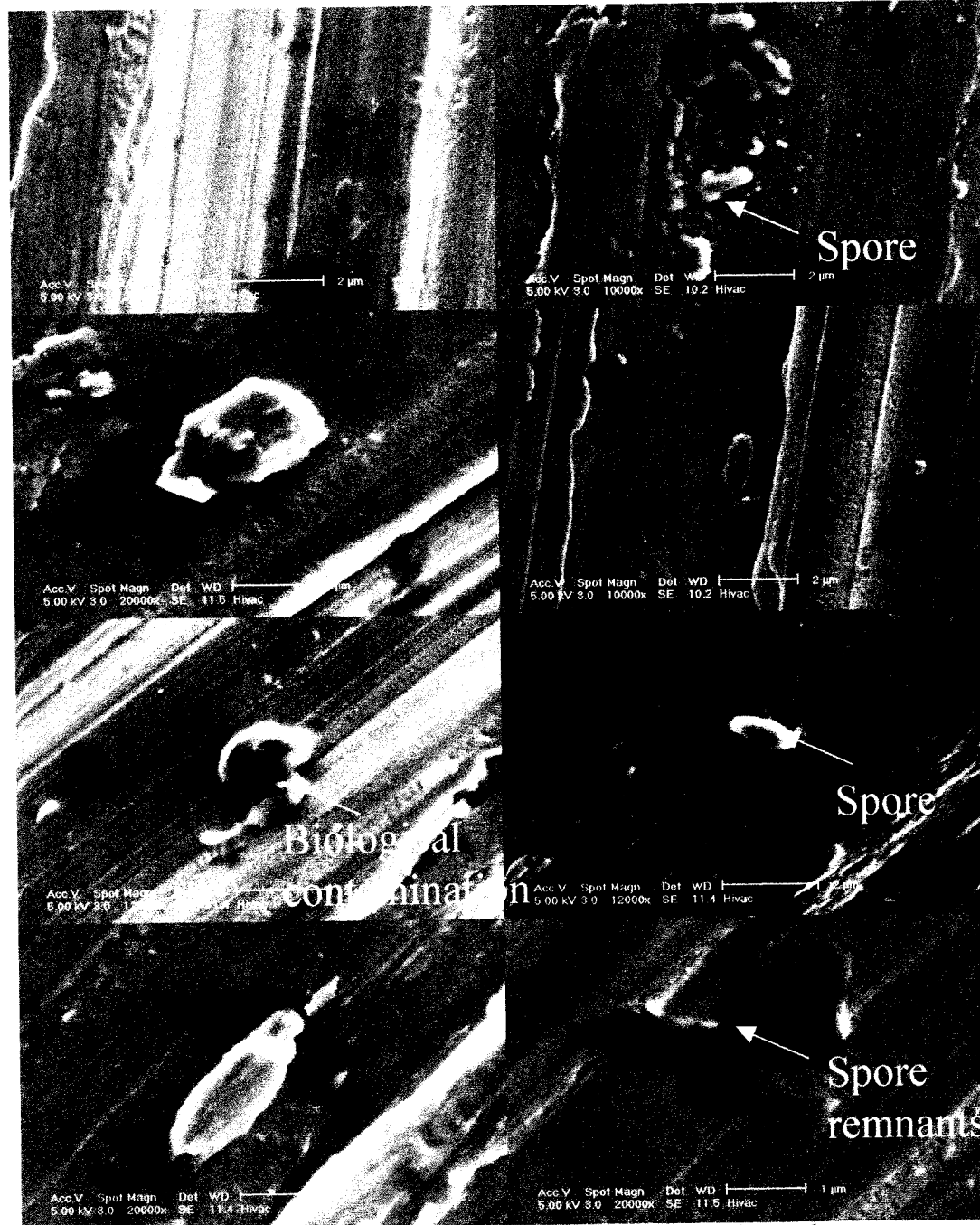
multiple solvent

detergent/  
ultrapure water

semi-aqueous  
multiple solvent

**Titanium  
6Al-4V  
(32 rms)**

ESEM



control

multiple solvent

detergent/  
ultrapure water

semi-aqueous  
multiple solvent

not inoculated

inoculated

**Titanium**  
**6Al-4V**  
**(32 rms)**

epifluorescence  
microscopy



Spore



control

multiple solvent

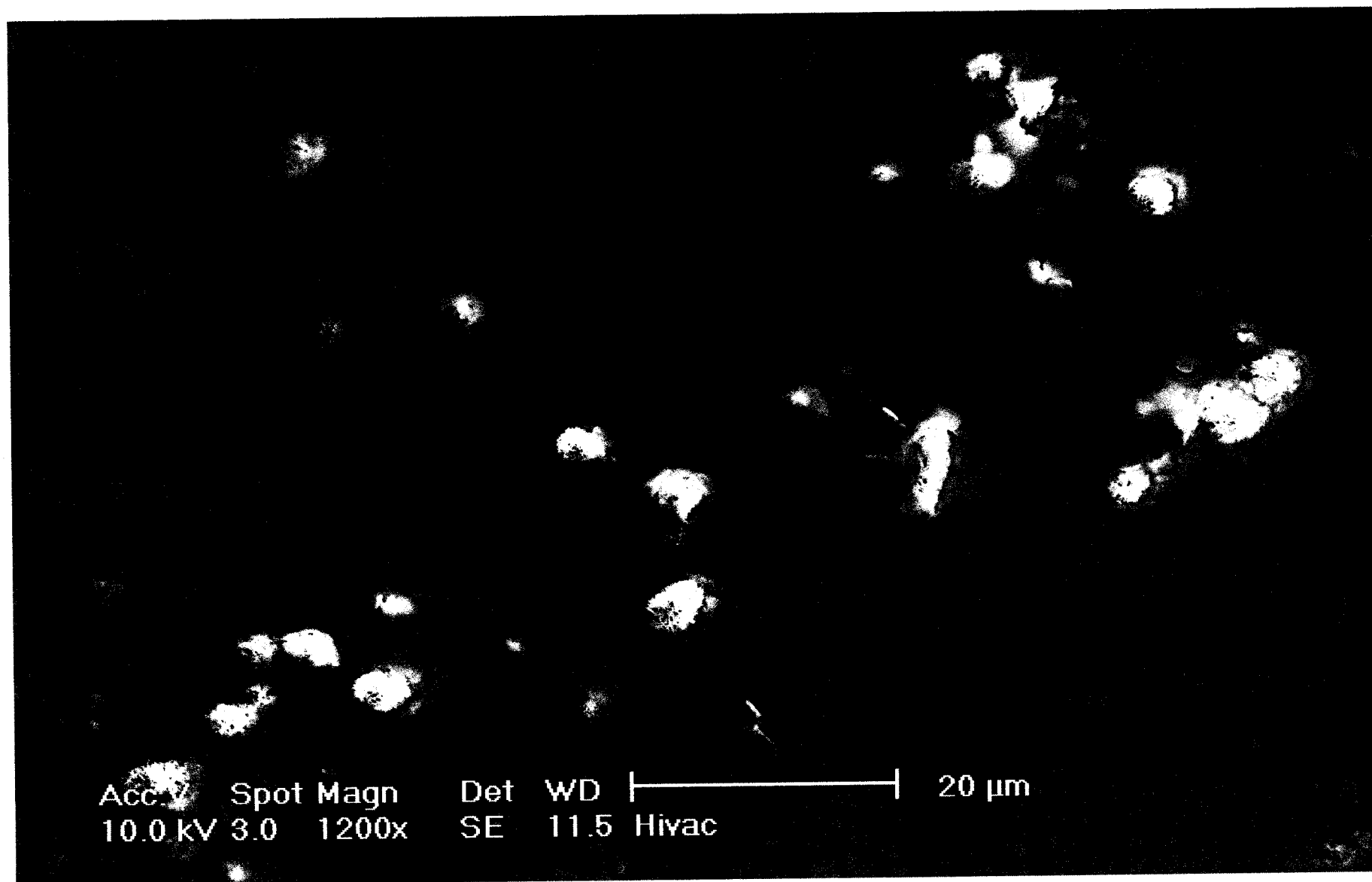
detergent/  
ultrapure water

semi-aqueous  
multiple solvent

coupon surface

filter collected

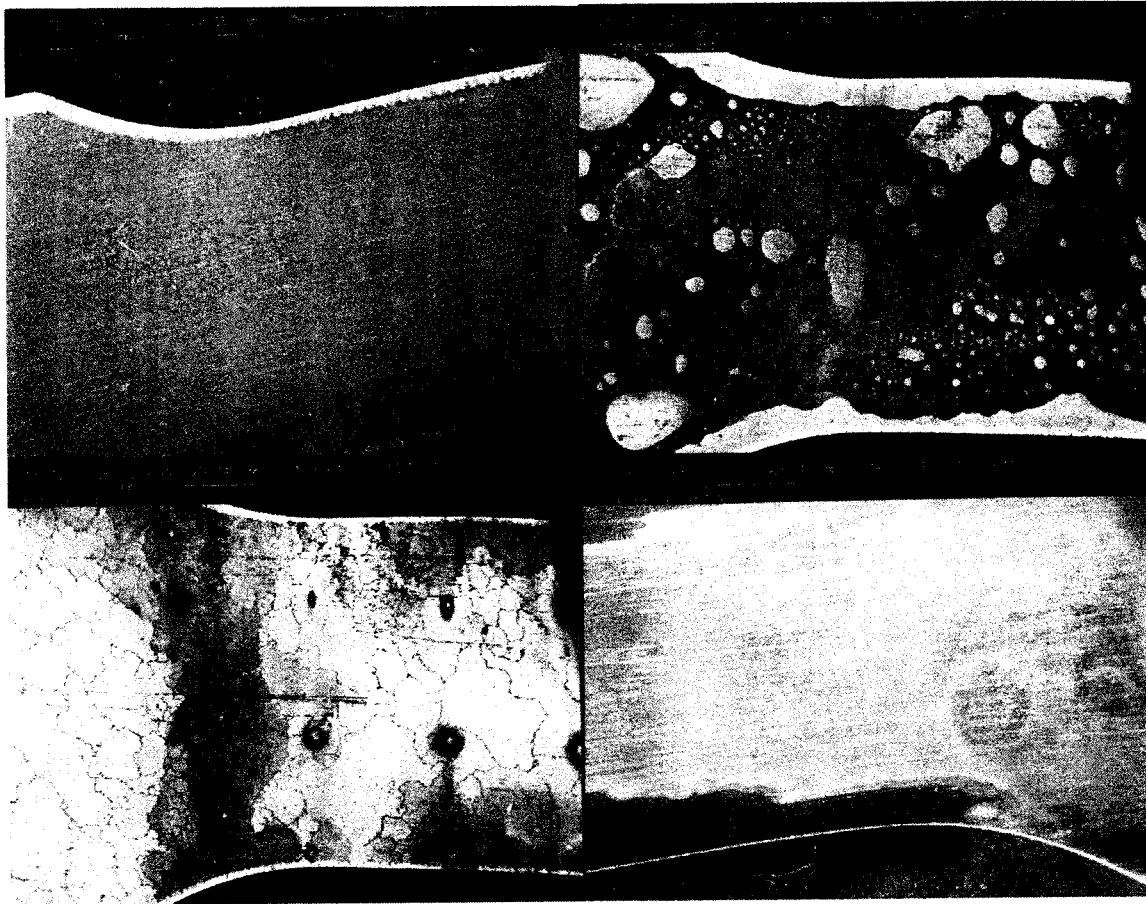
# Inoculated Al coupon cleaned by semi-aqueous, multiple solvent method



## Surface roughness/corrosion

Al 6061  
unpolished  
(15 rms)  
pre-cleaned

Al 6061  
unpolished  
(15 rms)  
pre-cleaned  
autoclaved and  
multiple solvent  
cleaned



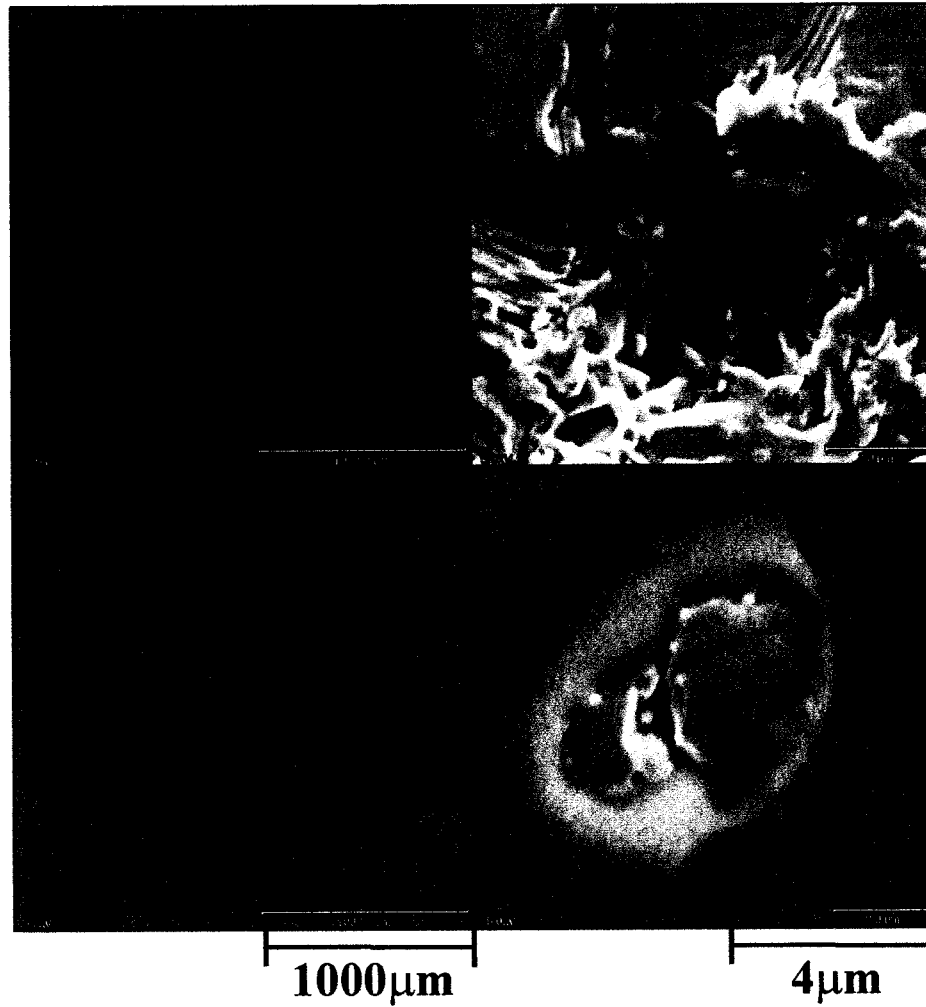
Al 6061  
unpolished  
(15 rms)  
pre-cleaned  
and autoclaved

Ti 6Al 4V  
pre-cleaned,  
autoclaved and  
multiple solvent  
cleaned

2 mm

polished Al 6061 after  
pre-cleaning with  
acetone, IPA and DCM

polished Al 6061 after  
pre-cleaning and  
autoclave sterilization



# Conclusions

**Cleaning matrix conducted as described using *B.subtilis* spores shows:**

- all cleaning methods studied clean to some degree
- Ti 6Al-4V (mill finish 32) can be “cleaned-to-sterility” using multiple-solvent cleaning methods (JPL method FS505146 Rev.C) with *no* spore remnants and only traces of organic contamination ( $\sim 50$  ng/cm<sup>2</sup>) remaining on the surface
- unpolished Al 6061 *cannot* be cleaned to sterility with any of the cleaning methods studied in the matrix
- polished Al 6061 easier to clean, but ability to “clean-to-sterility” needs to be confirmed with more replicate studies, i.e. may still be possible for spores to sit in 8 to 12 $\mu$ m surface defects seen on some polished Al 6061 coupons
- requisite autoclaving step leads to extensive oxidation of surface properties of hardware materials, thus results of cleaning matrix as conducted represent “worst case” cleaning scenario



- matrix leads to the formation of possible “spore houses” containing magnesium
- noted correlation between number of “spore houses” seen on coupons and vegetative growth
- number of possible “spore houses” containing magnesium found on polished Al about equal to number found on unpolished; however, size of “houses” on polished Al are smaller than those seen on unpolished Al, indicating “houses” may be composed of only inorganic materials

## Future Studies

- repeat clean-to-sterility studies on Ti and polished Al to confirm robustness of multiple solvent cleaning method
- compare surface properties of autoclaved Ti and Al surfaces to those found on aged flight hardware (e.g. Mars Pathfinder and Viking mission hardware)
- conduct more detailed chemical analyses of possible “spore houses”
- extend matrix to include composite hardware materials and vegetative microbes
- modify JPL multiple solvent method to include nitric acid passivation step for Al